

# Harmonizing Cerebrospinal Fluid Analysis for Multiple Sclerosis Investigation: An Update from the hCAMI Subcommittee of the Canadian Society of Clinical Chemists (CSCC)



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LABORATORIES

Leaders in Laboratory Medicine

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### INTRODUCTION

- Cerebrospinal fluid (CSF) oligoclonal banding (OCB) analysis, specifically ≥2 CSF-specific OCBs, can substitute for dissemination in time criteria of multiple sclerosis (MS) diagnosis <sup>1</sup>
- Associated tests (e.g., CSF immunoglobulin G (IgG), CSF albumin) and calculated indices (e.g., albumin quotient, IgG index) can support clinical interpretation
- There is significant variability in processes and reporting practices across Canadian clinical laboratories<sup>2</sup>
- To address this issue, the Harmonized CSF Analysis for MS Investigation (hCAMI) subcommittee (clinical chemists and neurologist) of the CSCC Reference Interval Harmonization Working Group was formed

# **OBJECTIVE**

Establish recommendations for laboratory processes and reporting of CSF OCB and associated tests supporting MS diagnosis.

# **METHODS**

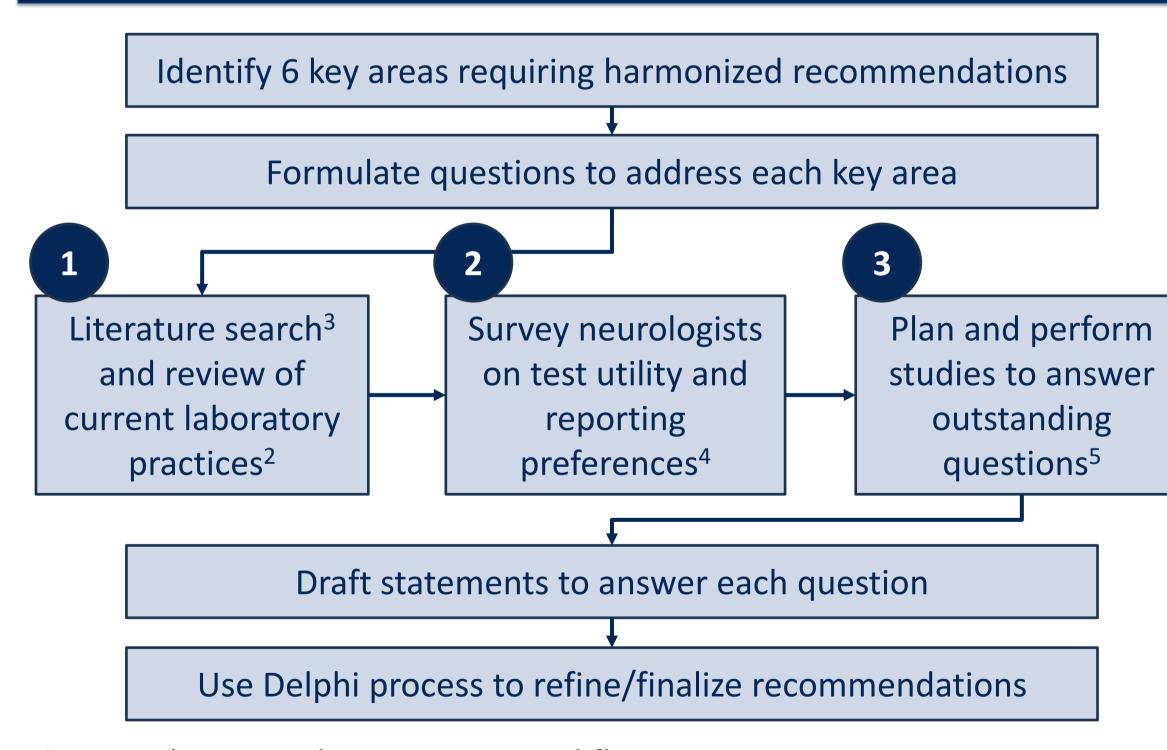
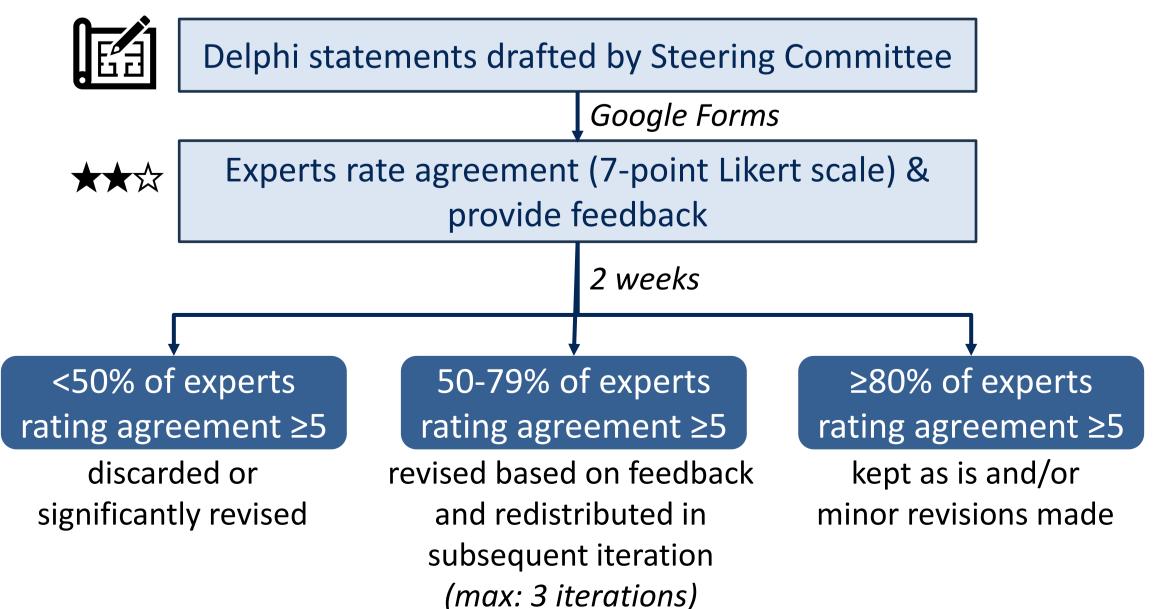
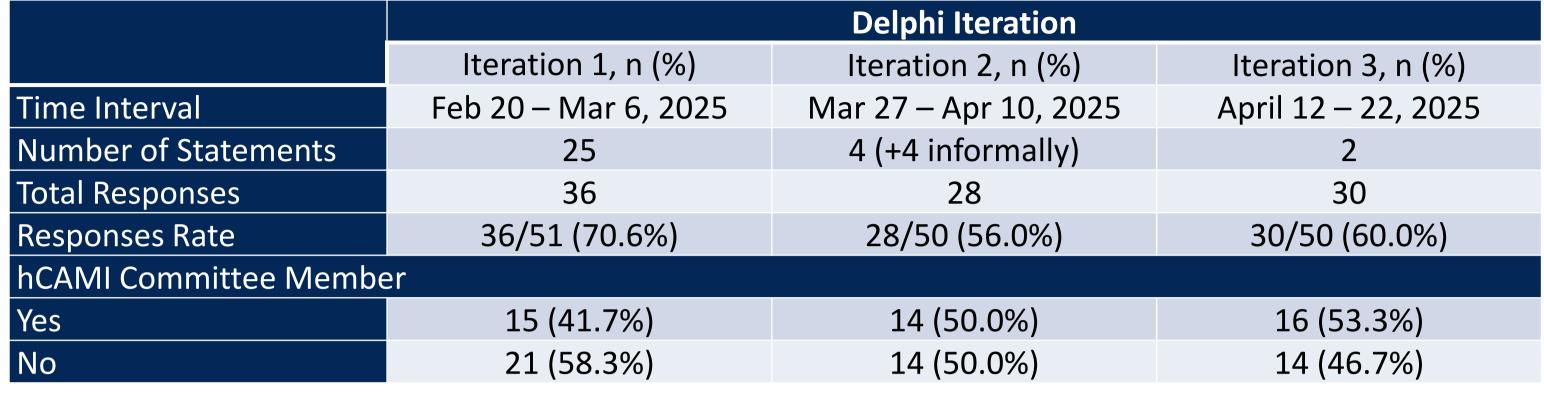


Figure 1. hCAMI Subcommittee Workflow



**Figure 2.** Modified Delphi Process. Town Hall held between Iterations 1 and 2 to provide evidence for statements that did not reach consensus.

 Table 1. Characteristics of each Delphi Iteration



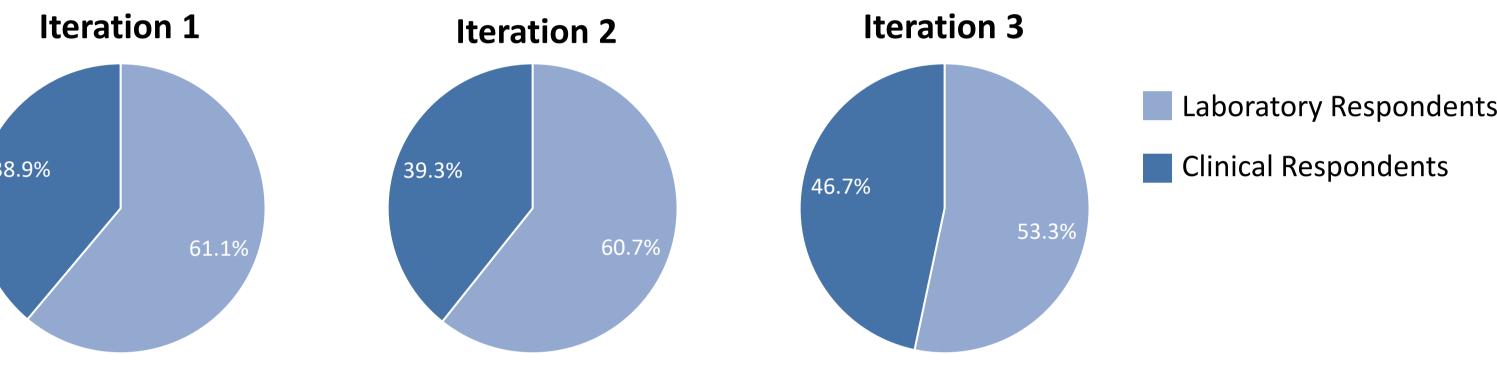


Figure 3. Subject Experts' Profession across all Iterations

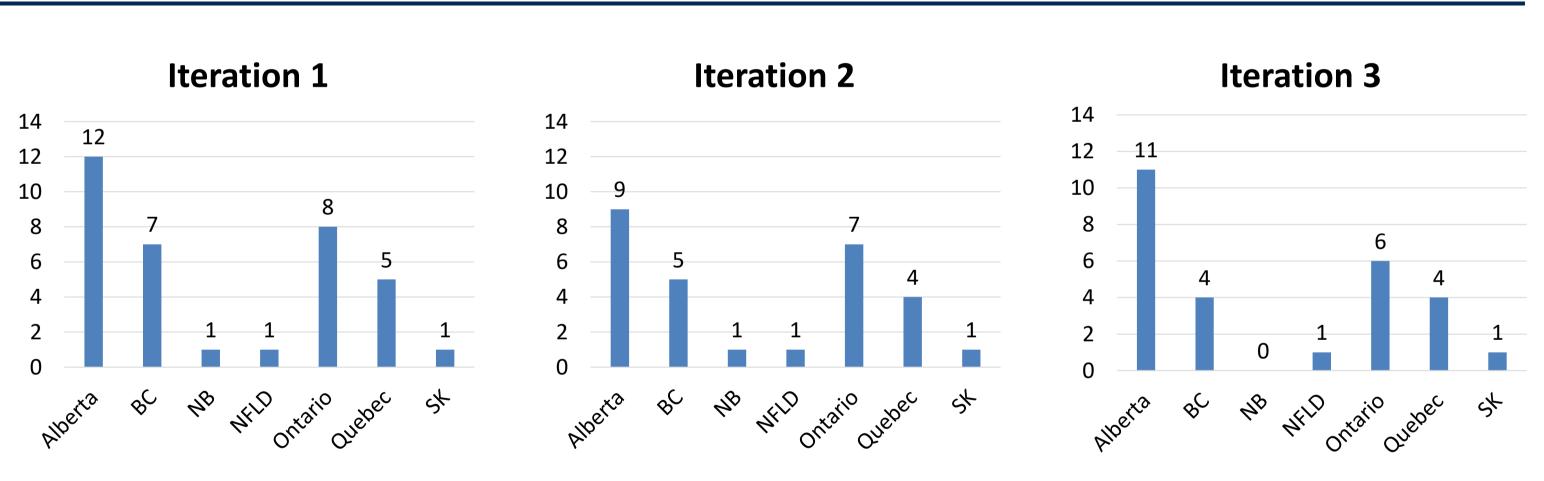


Figure 4. Subject Experts' Province of Practice across all Iterations

# CONCLUSION

- 24/25 Delphi Statements met consensus by Iteration 3 of the Delphi Process and will be formally recommended by the hCAMI subcommittee
- Recommendations to harmonize laboratory reporting will promote alignment of CSF OCB reporting practices with the latest evidence, ultimately enhancing diagnostic accuracy and patient care



RESULTS

Delphi Statement	Iteration		% Agreement ≥ 5 Overall (Lab/Clinical) Consensus criteria: ≥8
Section 1: Quality a	assurance practi	ces	
#1. QC material	1	35	89 (86/92)
#2. QC documentation	1	36	86 (96/71)
	2 (Informal)	28	93 (88/100)
#3. Competency assessment	1	36	97 (96/100)
#4. External quality assurance	1	36	97 (100/93)
#5. Unclear cases	1	35	63 (55/77)
	2	28	<b>75 (71/82)</b>
	3	30	100 (100/100)
Section 2: Plasma acceptability and time into	erval requireme	nts for paired CSF	and blood
#6. Plasma acceptability	1	35	97 (96/100)
#7. Paired specimen timing for CSF OCB	1	35	60 (52/71)
	2	28	96 (94/100)
#8. Paired specimen unavailable	1	36	83 (86/79)
	2 (Informal)	26	92 (94/90)
#9. Paired specimen unavailable & no CSF bands	1	36	92 (96/86)
#10. Paired specimen unavailable & 1 CSF band	1	36	94 (96/93)
#11. Paired specimen unavailable & 2+ CSF bands	1	35	94 (96/92)
#12. Paired specimen timing for associated tests	1	34	82 (70/100)
	2 (Informal)	27	89 (82/100)
Section 3: If and how to repo	ort CSF-specific	band counts	
#13. Reporting CSF-specific band counts	1	36	<b>75</b> ( <b>64</b> /93)
	2	28	75 (77/73)
	3	36	79 (80/79)
#14. Reporting CSF-specific band counts as categories	1	36	83 (96/64)
	2 (Informal)	26	77 (73/82)
#15. Reporting CSF-specific band counts using listed categories	1	35	74 (68/85)
	2	27	85 ( <mark>75</mark> /100)
Section 4: Interpretation and follow-up for mirror pattern	s (i.e., inflamma	atory response, m	onoclonal gammopath
#16. Mirror pattern	1	36	100 (100/100)
#17. Mirror pattern - monoclonal protein	1	36	89 (82/100)
#18. Mirror pattern - inflammatory response and/or monoclonal protei	in <sup>1</sup>	36	89 (82/100)
Section 5: Interpretation of matched bands w	ith differing inte	ensity between CS	F and serum
#19. Matched bands of differing intensity.	1	34	91 (91/92)
Section 6: Panel components and	reference interv	/als/decision_limit	S
#20. Panel components	1	34	97 (95/100)
#21. Decimal places & units	1	33	94 (90/100)
#22. Reference intervals	1	32	81 (80/83)
#23. Band count abnormal flagging	1	35	86 (86/85)
#24. Reference to MS clinical guidelines	1	35	89 (91/85)

# REFERENCES

- L. Olayinka, et al. Clin Chem Lab Med. (2025) Online ahead of print
- 2. A.J. Thompson, et al. *Lancet Neurol*. 17 (2) (2018) 162–173.
- 3. V. Higgins, et al. *Clin Biochem*. 116 (2023) 105–112.
- 4. V. Higgins, et al. Crit Rev Clin Lab Sci. (2025) 1–23. Online ahead of print
- 5. V. Higgins, et al. *Clin Biochem*. 135 (2025) 110855.